

**AMERICAN DENTAL ASSOCIATION
STATEMENT ON DENTAL UNIT WATERLINES***

Background: Organized dentistry has traditionally assumed responsibility for assessing and improving the quality of dental care provided to patients. The widespread adoption of enhanced infection control methodologies by dental practitioners is just one example of the profession's commitment to high quality patient care.

The Council is sensitive to heavy regulatory burden imposed on dentists in recent years by various federal, state and local government agencies. In some cases, the regulations have been based on limited science. The Council reaffirms its strong belief that both the profession and the public are served when recommendations affecting dental practice are based on sound science and take into account their cost in light of their expected benefit. The recommendations that follow are made in light of these considerations.

Through its continued monitoring of scientific literature, the Council has become aware that the microbiologic quality of water used in dental treatment could be improved. Although there is no evidence of a public health risk due to this phenomenon, steps should be taken to improve the quality of water used in patient care as soon as feasible. The profession, the dental industry, and the research community all have an important role to play in this process. Dental unit waterlines (the tubes that connect the high-speed handpiece, air/water syringe and ultrasonic scaler to the water supply) have been shown to harbor a wide variety of microorganisms including bacteria, fungi, and protozoans. These microorganisms colonize and replicate on the interior surfaces of the waterline tubing, inevitably resulting in adherent heterogenous microbial accumulations termed "biofilms." Biofilms, once formed, serve as a reservoir significantly amplifying the numbers of free-floating microorganisms in the water exiting the waterlines. It has been suggested that heating dental unit water to increase patient comfort, as is the practice in some dental offices, may further augment biofilm formation. In unmaintained dental unit waterline systems, these microbial accumulations can contribute to occasional objectionable odors and visible particles of biofilm material exiting the system.

Water Quality Improvement: Dental unit water systems currently designed for general dental practice are incapable of delivering water of an optimal microbiologic quality. The Council recommends an ambitious and aggressive course to encourage industry and the research community to improve the design of dental equipment so that by the year 2000, water delivered to patients during nonsurgical dental procedures consistently contains no more than 200 colony forming units per milliliter (cfu/ml) of aerobic mesophilic heterotrophic bacteria at any point in time in the unfiltered output of the dental unit; this is equivalent to an existing quality assurance standard for dialysate fluid that ensures the fluid delivery systems in hemodialysis units have not been colonized by indigenous waterborne organisms.

Manufacturers of dental equipment are encouraged to develop accessory components that can be retrofitted to dental units currently in use, whatever the water source (public or independent), to aid in achieving this goal. Further, the ADA should urge industry to ensure that all dental units manufactured and marketed in the U.S.A. in the future have the capability to be equipped with a separate water reservoir independent of the public water supply. In this way, dentists not only will have better control over the quality of the source water used in patient care, but also will be able to avoid interruptions in dental care when "boil water" notices are issued by local health authorities.

At the present time, commercially available options for improving dental unit water quality are limited and will involve some additional expense. They include the use of:

- o Independent water reservoirs
- o Chemical treatment regimens
- o Daily draining and air purging regimens
- o Point-of-use filters

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APPROVED
1996 MWM

*Adopted by the American Dental Association Board of Trustees, December 13, 1995 and ADA Council on Scientific Affairs, September 28, 1995

CHARACTERISTICS OF DUWL BIOFILM

1. Pioneer bacteria colonize new tubing wall with a "conditioning" layer
2. Biofilm grows in thickness and complexity
3. Sticky slime layer protects member organisms from physical and chemical removal and treatment
4. Consortium of participant microbes including bacteria, fungi, protozoa, and nematodes, depends on 90% water content within biofilm for nutrients
5. Water is stagnant 99% of the time allowing for release of organisms into water
6. Sloughing of pieces of dislodged biofilm provides gross downstream contamination
7. Biofilm sloughing is caused by manipulation of the tubing and flushing
8. Microbes protected by biofilm acquire enhanced resistance to chemical antimicrobials and sometimes antibiotics

| | | | |
|------------------------|-------------------|---------------------------------|--|
| Pasteurella spp. | water | opportunistic | wound infections, chronic respiratory infections |
| Proteus vulgaris | water | ? | urinary tract infection |
| Peptostreptococcus | mouth | ? | periodontal disease |
| Pseudomonas aeruginosa | water | opportunistic | septicemia, abscesses wound & respiratory infections |
| Pseudomonas cepacia | water | opportunistic | pneumonia, otitis, wound infections |
| Salmonella typhimurium | hands oral/ fecal | primary pathogen | diarrhea, septicemia |
| Streptococcus spp. | mouth | primary pathogen | respiratory infections endocarditis, meningitis |
| Xanthomonas | water | low | urinary tract infections, wound infections |
| FUNGI | | | |
| Penicillium | | allergenic / rare opportunistic | respiratory allergic reactions |
| Cladosporium | | allergenic / rare opportunistic | respiratory allergic reactions |
| Alternaria | | allergenic / rare opportunistic | respiratory allergic reactions |
| Scopulariopsis | | allergenic / rare opportunistic | respiratory allergic reactions |
| PROTOZOA | | | |
| Acanthamoeba | water | opportunistic | conjunctivitis, meningitis |
| Cryptosporidium | water | opportunistic | GI infection, severe dehydration |
| Microsporidium | water | opportunistic | GI infection |
| Giardia | water | opportunistic | diarrhea |

TABLE 3

| EFFECT OF IMMUNOCOMPROMISING CONDITIONS ON HOST RESISTANCE | |
|---|---|
| <u>CONDITION/FACTOR</u> | <u>SELECTED EFFECTS</u> |
| Neoplasia | Suppression of T-cell function; variable immunoglobulin levels |
| Nutritional deficiencies | Alteration of multiple immune mechanisms - epithelial integrity, phagocytosis, immunoglobulin synthesis |
| Aging | Decreased cellular immunity capabilities; decreased IgG responses to certain antigens |
| Alcoholism | Nutritional deficiencies; possible suppression of anti-bacterial inflammatory responses |
| Systemic Lupus Erythematosus (SLE) | Decreased delayed (Type IV) hypersensitivity; reduced complement activity |
| Asthma | Chronic bronchial inflammation; immediate hypersensitivity reactions to common inhalant allergens |
| Cystic Fibrosis | Susceptible to lower respiratory tract infections |
| Tuberculosis | Severe progressive TB may be associated with anergy (ie, lack of immune responsiveness) |
| Progressive HIV disease (AIDS) | Decreased CD ₄ -T-lymphocytes; decreased cellular immunity |
| Diabetes | Decreased phagocytic capability - diminished chemotaxis; poor bacterial ingestion |

Adapted from: Molinari, JA: *Compend Contin Ed Dent.* 16:130-132, 1995 and Ammann AJ: *Immunodeficiency Diseases*, in *Basic and Clinical Immunology* (6th edition), DP Stites, JD Stobo, JV Wells (eds), p. 317-355. Appleton & Lange Pub., Norwalk, 1987.

Diagnostic Quick Reference

| Signs and Symptoms | Environmental Tobacco Smoke PP-5 | Other Combustion Products PP-7 | Biological Pollutants pp-10 | Volatile Organics PP-13 | Heavy Metals PP-15 | Sick Bldg Syndrome pp-17 |
|--|-------------------------------------|-----------------------------------|--------------------------------|----------------------------|-----------------------|-----------------------------|
| Respiratory | | | | | | |
| Rhinitis, nasal congestion | ■ | ■ | ■ | ■ | | ■ |
| Epistaxis | | | | ■ ¹ | | |
| Pharyngitis, cough | ■ | ■ | ■ | ■ | | ■ |
| Wheezing, worsening asthma | ■ | ■ | | ■ | | ■ |
| Dyspnea | ■ ² | | ■ | | | ■ |
| Severe lung disease | | | | | | ■ ³ |
| Other | | | | | | |
| Conjunctival irritation | ■ | ■ | ■ | ■ | | ■ |
| Headache or dizziness | ■ | ■ | ■ | ■ | ■ | ■ |
| Lethargy, fatigue, malaise | | ■ ⁴ | ■ ⁵ | ■ | ■ | ■ |
| Nausea, vomiting, anorexia | | ■ ⁴ | ■ | ■ | ■ | |
| Cognitive impairment, personality change | | ■ ⁴ | | ■ | ■ | ■ |
| Rashes | | | ■ | ■ | ■ | |
| Fever, chills | | | ■ ⁶ | | ■ | |
| Tachycardia | | ■ ⁴ | | | ■ | |
| Retinal hemorrhage | | ■ ⁴ | | | | |
| Myalgia | | | | ■ ⁵ | | ■ |
| Hearing loss | | | | ■ | | |

¹Associated especially with formaldehyde. ²In asthma. ³Hypersensitivity pneumonitis, Legionnaires' Disease. ⁴Particularly associated with high CO levels. ⁵Hypersensitivity pneumonitis, humidifier fever. ⁶With marked hypersensitivity reactions and Legionnaires' Disease.

Particular Effects Seen in Infants and Children

Environmental Tobacco Smoke: frequent upper respiratory infections, otitis media; persistent middle-ear effusion; asthma onset, increased severity; recurrent pneumonia, bronchitis.

Acute Lead Toxicity: irritability, abdominal pain, ataxia, seizures, loss of consciousness.

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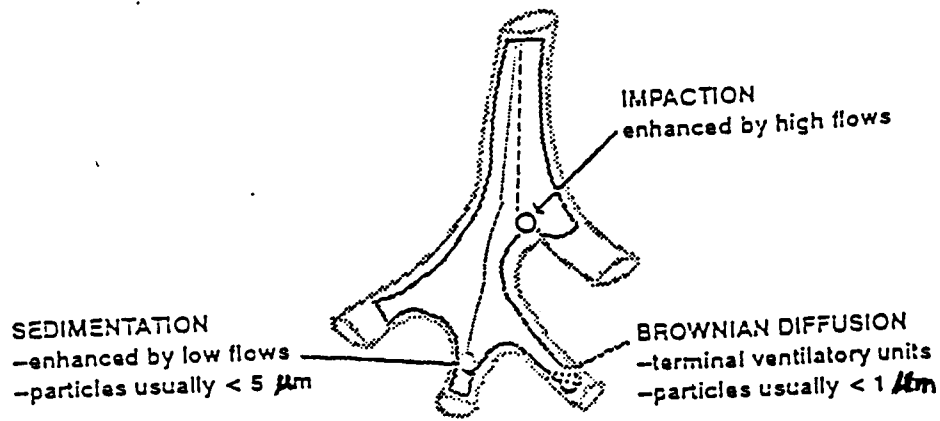


FIG. 1. Mechanisms of aerosol deposition in lung. (Modified from Murray JF, Nadel JA, eds. Textbook of respiratory medicine. Vol 2. Philadelphia: WB Saunders, 1988:313-31.)

TABLE I. Typical sizes for some common aerosol particles

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| <ul style="list-style-type: none"> • Tobacco smoke: $< 1 \mu\text{m}$ • Pollens: Approximately 10-30 μm • Mold spores: Approximately 2-50 μm • Actinomycete spores: Approximately 1-2 μm • Foundry dusts: Approximately 50 μm |
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TABLE II. Major factors determining particle deposition

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| <ul style="list-style-type: none"> • Respiratory tract anatomy • Patterns of airflow • Mode of inhalation • Particle characteristics (size, density, charge) |
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J ALLERGY CLIN IMMUNOL 1994;94:304-9.
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0091-6749/94 \$3.00 + 0 1/0/56009